



Azma Elixir Pajooh

DNrich FFPE Kit

Cat. No. AEFDX-1059

For Research Use Only

Introduction

The **DNrich** FFPE Tissue Kit provides all of the reagents necessary to extract DNA from a wide variety of Formalin-Fixed Paraffin Embedded tissue sources. DNA purified with this kit is suitable for a variety of applications, including amplification and digestion with restriction endonucleases.

DNrich FFPE Tissue Kit components*

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G Solution	50 ml
ZR Solution(conc.) **	15 ml
TD Buffer	20 ml
Activator Reagent**	2 ml
VD Buffer	10 ml
SE Buffer	15 ml
Wash Buffer(conc.) **	15 ml
Elution Buffer	3 ml
Column	50
Manual	1

* 1.5 ml microcentrifuge tube, **molecular biology grade water** and **absolute ethanol** are needed but are not included.

** Please refer to reminder, **Activator Reagent, Wash Buffer and ZR Solution Preparation** before using this kit

Chemical Hazard

Always wear gloves and practice standard safety precautions while using the kit. Do NOT disinfect extraction waste in solutions containing **bleach** or any other form of acid. To clean any items contaminated with the reagent, simply soak in detergent and water to remove all traces of contamination before cleaning with bleach or acidic solutions.

Reminder

- Pre-set heater block at 65°C.
- Prepare **Activator Reagent** immediately prior to use. Prepared Activator Reagent must be kept at 4°C.
- Prepare **Wash Buffer** before first use.
- During DNA extraction, **never** open the microtube cap outside the laminar hood.

Activator Reagent Preparation

- Add **2 ml** of **molecular** biology grade water to **Activator Reagent**, and vortex it well.

Note: Mark the check box on the bottle and write the date.








Note: For the best results, the prepared Activator Reagent should be used immediately. Prepared Activator Reagent can be stored for up to 3 months or 6 months at 4°C and -20°C, respectively.

Wash Buffer and ZR Solution Preparation

- Add **35 ml molecular grade absolute ethanol** to **Wash Buffer** and **ZR Solution** bottle before first use and mark the check box on the bottle.

Symbols

The following symbols are used in the labeling:

	Catalogue number
	Manufacturer
	Temperature limit
	Batch code
	Use-by date
	Contains sufficient for <n> tests
	Consult instructions for use

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PROTOCOL

Step 1: Sample Preparation

- Transfer **50 mg** (up to 200 mg) of tissue sample into a 1.5 ml tube.

Step 2: Paraffin Removal

- Add **500 µl G Solution** to sample tube and vortex vigorously.
- Incubate at **65°C** for **2 minutes**.
- Centrifuge at **12000 g** for **5 minutes**.
- **Decant** the supernatant into waste.
- **Repeat step 2.**

Step 3: Tissue Washing

- Add **500 µl ZR Solution** to sample tube and vortex it.
- Centrifuge at **12000 g** for **5 minutes**.
- **Decant** the supernatant into waste.
- **Note:** Repeat this step to **extract a pure DNA**.

Step 4: Tissue Digestion & Lysis

Note: Refer to reminder before use activator reagent

- Add **400 µl of TD Buffer** and **40 µl of Activator Reagent** to the sample tube and vortex vigorously.
- Incubate at **65°C** for **30 minutes**.
Note: During incubation time, vortex the sample tube every 10 minutes.
- Incubate at **85°C** for **10 minutes** and vortex every 5 minutes.
- Add **200 µl of VD Buffer** to sample tube and vortex vigorously and then keep at RT for **5 minutes**.
- Centrifuge at **12000 g** for **10 minutes**.
Note: Do not disturb the phases.
- Carefully transfer about **300µl of supernatant** to a new 1.5 ml tube.
- Add **300µl of SE Buffer** to the tube, invert for 5 times and keep at room temperature for **3 minutes** and then transfer all the sample to a **spin column**.
- Centrifuge at **2000 g** for **2 minutes** and **discard** the flow through and reassemble the spin column with its collection tube.

Step 5: Washing

- Add **500 µl of Wash Buffer** to the column, centrifuge at **8000 g** for **1 minutes** and **discard** the flow through and reassemble the spin column with its collection tube.
- **Repeat step 5.**

Step 6: Column Drying

- Centrifuge at **8000 g** for **1 minute**.
- **Discard** the collection tube and transfer the spin column to a new **1.5 ml** microcentrifuge tube.

Step 7: DNA Elution

- Add **50 µl of Elution Buffer** to the center of column and let stay at **RT** for **3 minutes**.
- Centrifuge at **10000 g** for **2 minutes**.